

Concomitant Use of Analgesics and Immune Checkpoint Inhibitors in Non-Small Cell Lung Cancer: A Pharmacodynamics Perspective

By Rahmad Aji Prasetya

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ABSTRACT

The invention of immunotherapy, such as immune checkpoint inhibitors (ICIs) for advanced-stage non-small cell lung cancer (NSCLC), has become a new standard of care for a defined group of NSCLC patients. However, the possible impacts of ICI interactions with analgesics for alleviating cancer-related pain are unclear and lack clinical evidence. Many studies have indicated that opioids detrimentally affect the immune system, possibly harming patients of ongoing immunotherapy. Opioids may repress the immune system in various ways, including impairing T cell function, upregulating immunosuppressor Treg cells, and interrupting intestinal microflora composition that disrupts the entire immune system. Furthermore, opioids can influence tumor progression and metastasis directly as opioid receptors are overexpressed in several types of NSCLC. In contrast, another analgesic acting on cyclooxygenase (COX) inhibition (i.e., NSAIDs) may be a candidate for adjuvant therapy since COX-2 is also expressed in the tumor cells of NSCLC patients. In addition, COX-2 is associated with tumor proliferation and metastasis. Therefore, both prospective and retrospective studies should confirm the advantages and disadvantages of the concurrent use of analgesics and ICIs in a clinical setting.

Keywords: immune checkpoint blockers, NSAIDs, opioids, T-lymphocyte, gut microbiota

1. Introduction

Lung cancer is the primary cause of cancer mortality in men and the second-highest contributor to cancer mortality (after breast cancer) in women globally. In 2018, the World Health Organization (WHO) estimated 2.09 million cases and 1.76 million deaths caused by this cancer (Torre et al., 2016; WHO, 2018). Among these cases, non-small cell lung carcinomas (NSCLC) accounted for 85%-90% of all lung cancer types (Lemjabbar-Alaoui et al., 2015). Treatment options for NSCLC vary, including surgery, radiotherapy, platinum-based chemotherapy, targeted therapy, and immunotherapy, depending on cancer stages, histologic classification, and genetic alteration. Most NSCLC patients (>70%) are diagnosed during stages III and IV (Cheema et al., 2019), and platinum-based chemotherapies have become standard care, although their clinical efficacy is restrained by hemato- and neuro-toxicities (Hirsch et al., 2017; Lemjabbar-Alaoui et al., 2015). Recently, molecularly targeted agents, such as erlotinib and gefitinib, which specifically block epidermal growth factor receptors (EGFRs), have been indicated as the first-line treatment for patients who carry an EGFR mutation in their tumors. However, these agents cover only 10%-17% of the NSCLC patient population (Hirsch et al., 2017; Langer, 2015). Meanwhile, 31% of NSCLC patients' tumor oncogenic drivers remain undetected. The availability of suitable drugs for patients lacking a detectable gene mutation presents a drawback to this targeted treatment (Hirsch et al., 2017; Mayekar and Bivona, 2017). Furthermore, improvement in lung cancer treatment has plateaued over the last ten years. Hence, a novel strategy is needed for broader applicability to encounter another pathophysiology of tumor evasion from immunosurveillance.

Although tumors express antigens recognizable by the immune system, they possess poor immunogenicity. Hence, antigen presentation alone is inadequate in activating T cells. Besides the binding of the T cell receptor (TCR) to the antigenic peptide bound to the major histocompatibility complex (MHC) of the antigen-presenting cell (APC), the additional stimulatory co-signal provided by co-receptors is necessary. These co-receptors are crucial for regulating T cell response and balancing co-stimulatory and inhibitory (i.e., immune checkpoint) signals. Cancerous cells utilize several strategies to evade immune checkpoints regulated by T cells, enabling them to grow and spread unchecked (Pardoll, 2012). These strategies provide a negative immune checkpoint signal to T cells, which is specific against tumor antigens. Thus, anticancer responses from the immune system could be repressed. Monoclonal antibodies have been indicated as a blockade to immune checkpoint pathways, including cytotoxic T-lymphocyte-associated antigen 4 (CTLA-4), programmed death protein 1 (PD-1), and its ligands, PD-L1 (Abdel Karim and Kelly, 2019; Langer, 2015; Memon and Patel, 2019). To date, only PD-1 (pembrolizumab and nivolumab) and PD-L1 (atezolizumab and

durvalumab) have been used for NSCLC patients in a clinical setting after gaining Food and Drug Administration (FDA) approval, as first-line monotherapy, in combination, or as second-line therapy after chemotherapy (Abdel Karim and Kelly, 2019; Brahmer et al., 2017). Thus, immune checkpoint inhibitors (ICIs) have continued to expand as an active field of research in advanced NSCLC.

In clinical practice, challenges in treating cancer patients rely on eradicating the tumor and addressing the symptoms while focusing on the current medication's adverse events (Simoff et al., 2013; Temel et al., 2006; Walling et al., 2015). Pain, the most distressing symptom causing depression and anxiety, is experienced by 58.2% of advanced-stage lung cancer patients (Walling et al., 2015). Hence, analgesic use is critical but generates new challenges for drug interactions due to concomitant medications. Therefore, this review provides information on (i) the pharmacological mechanism in which analgesics interfere with the therapeutic efficacy of the ICI specific for NSCLC, and (ii) the best type of analgesics, including dosage regimentation when used concomitantly. The ICI here is limited to the drugs acting on CTLA-4 and PD-1 pathways as they have been widely used in clinical settings and trials.

2. Pharmacodynamics of Immune Checkpoint Inhibitors

Under non-pathologic conditions, immune checkpoints are essential to prevent autoimmunity and protect tissue damage from excessive inflammation when the immune system neutralizes pathogenic infections. Moreover, tumor cells can dysregulate immune checkpoint ligands and receptors, resulting in overexpression in the cell itself and surrounded non-transformed cells in the tumor microenvironment. Therefore, ICIs target the receptors or ligands expressed on the lymphocyte to augment antitumor activity (Pardoll, 2012).

This section first discusses the pharmacodynamics of ICIs before elaborating on the analgesic interference mechanism. The concomitant use of ICIs and analgesics is mainly interconnected in the immune system. Thus, each drug's adverse effects are highlighted to determine if side effects increase when a drug is combined simultaneously.

2.1 Blockage of CTLA-4 in the early phase of T cell activation

CTLA-4 was the first clinically targeted inhibitory T cell co-receptor because the antagonist CTLA-4 antibody could induce therapeutic antitumor immunity in the mouse model in 1996 (Leach et al., 1996). Essentially, CTLA-4 is a counterbalance of CD28, a co-stimulatory receptor of T cell. T cell activation mostly requires CD28 colligation, in addition to the binding of TCR to the peptide antigen,

for stabilizing messenger RNA (mRNA) of cytokines and inducing the activation of the nuclear factor κ B (NF- κ B) and the nuclear factor of activated T cells (NFAT). CD28 is expressed and localized on the surface of naive and activated T cells, while CTLA-4 is primarily located in intracellular compartments and expressed in response to TCR/CD28 co-stimulation at the early activation stages. Upon TCR ligation, calcium influx and the TCR-interacting molecule (TRIM) control the release of CTLA-4 from the trans-Golgi network (TGN) or lysosome or endosome to the cell surface (Rudd et al., 2009).

To our knowledge, the intrinsic signaling in which CTLA-4 dampens T cells' immune response has been controversially discussed with no consensus of the distinct signaling pathway of CTLA-4 (Mitsuiki et al., 2019; Walker and Sansom, 2015). However, it is widely accepted that CTLA-4 and CD28 bind to the same ligands CD80 (B7-1) and CD86 (B7-2). The overall binding affinity is higher for CTLA-4 than CD28, so that the intrinsic inhibitory signal will rise after CTLA-4 outcompetes the binding of CD28, whereas CTLA-4 is activated later than CD28 (Figure 1). Furthermore, other concepts of CTLA-4 functions have been proposed and discussed, including a magnification of regulatory T (Treg) cell immunosuppressive activity (Gardner et al., 2014; Leach et al., 1996; Pardoll, 2012; Rudd et al., 2009). Specifically, in NSCLC patients, Erfani et al. (2012) stated that CTLA-4 molecules found in the intracellular compartment were higher than presented on the surface of lymphocytes subsets. However, only surface CTLA-4 on the CD8⁺ lymphocytes was significantly higher than control healthy subjects, which was inconsistent with the regulation of CTLA-4 expression. Nevertheless, CTLA-4 is rapidly internalized immediately after activation. Thus, this internalization would also serve as an inhibitory mechanism due to the TCR-antigen complex's co-internalization resulting in the unresponsiveness of lymphocytes (Erfani et al., 2012).

Anti-CTLA-4 agents like ipilimumab and tremelimumab have received FDA approval to treat advanced melanoma and mesothelioma, respectively, but not for lung cancer. Moreover, the clinicaltrials.gov database lists at least 16 active phase-3,4 clinical trials for NSCLC treatment using anti-CTLA-4, such as ipilimumab and tremelimumab. Among these trials, anti-CTLA-4 is mostly used in combination with anti-PD-1 (e.g., pembrolizumab, nivolumab, REGN2810) or PD-L1 (durvalumab), as shown in Table 1 (US National Library of Medicine, 2015). Many studies used drug combination of ipilimumab and nivolumab, and one of them has completed results (NCT02477826). It showed that among patients with a tumor mutational burden of at least ten mutations per megabase, those who received nivolumab plus ipilimumab have significantly longer progression-free survival (PFS) than patients receiving platinum doublet chemotherapy (Hellmann et al., 2018). Regarding the safety profile of this combination, the incidence of treatment-related adverse events was similar compared

to chemotherapy. Then, the most common adverse events were related to skin reactions (33.9%); (Hellmann et al., 2018).

2.2 Anti PD-1/PD-L1: The more specific the target, the more potent the activity

PD-1, like CTLA-4, is a member of the co-inhibitory receptor that serves as an immune checkpoint. In contrast to CTLA-4, which has a significant role in the early stage of T cell activation, PD-1 mainly expresses on cells within the tumor microenvironment to limit inflammatory response and autoimmunity in later stages. PD-1-known ligands, PD-L1 (B7-H1 or CD274) and PD-L2 (B7-DC or CD273), are inhibitory. However, PD-L1 expression is more prevalent than PD-L2 in various cancers, hence becoming a target for immunotherapy. PD-1/PD-L1 interaction is activated by several potent inducers, such as IFN- γ and TNF- α , produced by activated type 1 T cells. Meanwhile, GM-CSF and VEGF are secreted by a variety of cancer stromal cells (Figure 2a), and PD-L1 is upregulated by oncogenes (inherent immune resistance) so that cancer cells can overexpress PD-L1 (He et al., 2015; Pardoll, 2012; Figure 2b). D’Incecco et al. (2015) confirmed this finding in their study of PD-1/PD-L1 expression in tumor cells of NSCLC patients, revealing that PD-1 expression was mostly presented in the patients with KRAS mutations, while the ligand PD-L1 was significantly associated with the presence of EGFR mutations or ALK translocations (D’Incecco et al., 2015). Comparing PD-1/PD-L1 expression in NSCLC patients and healthy donors, Arrieta et al. (2017) and Meniawy et al. (2016) observed peripheral blood mononuclear cells (PBMCs) from both subjects. They found that NSCLC patients had a significantly higher proportion of PD-L1 in circulating CD3⁺T lymphocyte and CD3⁺CD8⁺ cytotoxic T-lymphocyte cells than healthy donors (Arrieta et al., 2017; Meniawy et al., 2016). PD-1/PD-L1 ligation affects the immune system’s suppression through multiple mechanisms, such as inducing apoptosis of activated T cells, facilitating T cell energy and exhaustion, and enhancing Treg cell function (He et al., 2015).

National Comprehensive Cancer Network Clinical Practice Guidelines in Oncology (NCCN Guidelines) for Non-Small Cell Lung Cancer V.2.2020 include all FDA-approved anti-PD-1/PD-L1 in the therapy recommendation for advanced or metastatic NSCLC with PD-L1 expression \geq 1% (guideline code NSCL-28/29), except for durvalumab. Durvalumab is proposed as a consolidation treatment following the absence of disease progression after radiation therapy regardless of the cancer type or stage (guideline code NSCL-E/3/6/7/9/12/13/). Among these immunotherapies, pembrolizumab (Keytruda®) is the only assigned single agent for the first-line treatment of patients expressing PD-L1 \geq 50%, and even \geq 1%-49% expression can be considered for patients with poor performance status or contraindicated to combination chemotherapy (National Comprehensive Cancer Network, 2019). Besides, many ongoing phase-3/4 clinical trials investigate the efficacy and safety of PD-1 or PD-L1

either as monotherapy or in combination with chemotherapy, kinase inhibitors, or monoclonal antibody targetting TIGIT receptor (Table 1).

Thus, the ICI works mainly to manipulate the endogenous antitumor activity of T lymphocyte subsets, mainly by blocking the inhibitory co-receptors that affect downstream signaling. T cells become the main focus of efforts to regain their immune capacity to (1) selectively distinguish cancerous antigens from their normal counterparts, (2) directly kill cancer cells, and (3) govern the diverse immune response that integrates many other cells, including dendritic cells, B lymphocytes, NK cells, and others (Pardoll, 2012). Since ICI therapy's success relies on patients' immunity response, other substances that intervene in the immune system could hamper ICI effectiveness, such as drugs to alleviate symptoms like pain.

1 Table 1. Ongoing clinical trials phase 3/4 of immune checkpoint inhibitors for NSCLC (The data was compiled from clinicaltrials.gov
 2 and accessed on April 2021)

ICI agents	Treatment Regimen	Clinical Trial Identifier
Anti-CTLA-4		
Ipilimumab	Ipilimumab+Nivolumab	NCT02477826, NCT02998528, NCT03351361, NCT03469960, NCT03391869, NCT02869789
	Ipilimumab+Nivolumab+Chemoradiotherapy	NCT04026412, NCT03215706
	Ipilimumab+Pembrolizumab	NCT03302234
	Ipilimumab+REGN2810	NCT03515629
	Ipilimumab+REGN2810 with Chemotherapy	NCT03409614
Tremelimumab	Tremelimumab+Durvalumab	NCT02453282, NCT02542293
	Tremelimumab+Durvalumab+Chemotherapy	NCT03164616
Anti-PD-1		
Nivolumab	Nivolumab (monotherapy)	NCT03542461, NCT02066636, NCT02477826, NCT01673867, NCT01642004, NCT03195491, NCT02613507, NCT02595944, NCT02713867, NCT04157985, NCT02041533
	Nivolumab+Chemoradiotherapy	NCT04026412
	Nivolumab+Chemotherapy	NCT02477826, NCT02864251, NCT02998528, NCT04025879, NCT04564157
	Nivolumab+ Sitravatinib	NCT03906071
Pembrolizumab	Pembrolizumab (monotherapy)	NCT03134456, NCT04676412, NCT04738487, NCT03715205, NCT03867175, NCT04475939
	Pembrolizumab+Chemotherapy	NCT04547504, NCT03425643, NCT03774732, NCT04267848, NCT04222972, NCT04194944, NCT03793179
	Pembrolizumab+Lenvatinib	NCT04676412, NCT03976375
	Pembrolizumab+Radiation	NCT03924869, NCT03867175
	Pembrolizumab+Chemoradiation	NCT04380636
	Pembrolizumab+Chemotherapy+Lenvatinib	NCT04716933
	Pembrolizumab+Carboplatin+Taxane+ Olaparib	NCT03976362, NCT03976323

Pembrolizumab+Chemotherapy+ Radiotherapy	NCT03774732
Pembrolizumab+Niraparib	NCT04475939
Anti-PD-L1	
Durvalumab (monotherapy)	NCT04381494, NCT04513925, NCT04642469, NCT03706690, NCT04078152
Durvalumab+Chemotherapy	NCT03800134, NCT04385368, NCT04092283
Durvalumab+Chemoradiotherapy	NCT04026412, NCT04380636, NCT01993810
Durvalumab+ Tremelimumab+Chemotherapy	NCT03164616
Durvalumab+Radiotherapy	NCT03833154
Atezolizumab (monotherapy)	NCT03991403, NCT03735121, NCT03178552
Atezolizumab+Tiragolumab	NCT04513925, NCT04294810
Atezolizumab+Chemotherapy	NCT03977194, NCT03456063
Atezolizumab+Radiotherapy	NCT04214262
Atezolizumab+Bevacizumab+Chemotherapy	NCT04194203
Atezolizumab+Cabozantinib	NCT04471428

4 **3. Analgesics Affecting the Immune System**

5 The chance of drug interaction occurring between ICIs and analgesics is high, as Simone et al. (2012)
6 reported that 92% of lung cancer patients experience pain either due to the malignancy or the
7 treatment (Simone et al., 2012). Moreover, a retrospective study of analgesic prescription patterns in
8 over 10,000 lung cancer patients showed that approximately 55% were prescribed level 3 analgesics,
9 including morphine, diamorphine, fentanyl, alfentanil. Meanwhile, another half of the patients
10 received weaker opioids (e.g., codeine, tramadol) or non-steroidal anti-inflammatory drugs (NSAIDs).
11 This study recorded the prescription of these analgesics in the last three months of lung cancer
12 patients' lives (Gao et al., 2011). Although the information on the cancer type and previous/current
13 cancer therapy were unavailable, this study demonstrated that at least half the patients received
14 opioids for any purpose.

15

16 Opioids are potent analgesics for severe pain, yet they are accompanied by many non-analgesic
17 effects ranging from constipation to respiratory depression. Another potential risk is immune
18 response suppression, which could be a drawback for immune-related anticancer treatment. The
19 crosstalk between opioids and anticancer immunity has been investigated widely, with varying
20 results depending on drug type and duration of use (Liang et al., 2016). Unlike opioids that may
21 repress the immune system, NSAIDs, particularly selective cyclooxygenase-2 (COX-2) inhibitors, may
22 corroborate cancer therapy's response rate or at least not contraindicate it (Yokouchi and Kanazawa,
23 2015). Hence, this section discusses the effects of analgesics, both opioids, and NSAIDs, within the
24 immune system, particularly which potentially interrupt the mechanism of ICI pharmacodynamics,
25 either directly or indirectly.

26

27 3.1 Opioid-mediated adaptive immune compromise

28 Many studies have shown that chronic opioid exposure is associated with the shorter survival of
29 NSCLC patients, although conclusions have been drawn from retrospective studies of morphine use
30 in postoperative settings as palliative care or before chemotherapy initiation (Hasegawa et al., 2018;
31 Wang et al., 2015; Zylla et al., 2014). Recent studies have emerged that identify opioids'
32 immunosuppressive characteristics, causing NSCLC patients' poor survival. Review articles written by
33 Liang et al. (Liang et al., 2016), Plein and Rittner (Plein and Rittner, 2018), Roy et al. (Roy et al., 2011),
34 and Zajączkowska et al. (Zajączkowska et al., 2018) have comprehensively presented the mechanism
35 of how opioids affect both innate and adaptive immunity.

36

37 Because of ICIs' modulation to T lymphocyte subsets, we examine the effect of opioids, especially
38 morphine, on T cells and their relation to the adaptive immune system. Morphine is the first-line

39 opioid employed to manage cancer-related pain. Morphine's effects are typically mediated by
40 specific opioid receptors (μ , δ , and κ) either in the central or peripheral nervous system. Outside the
41 nervous system, opioid receptors are also found in T lymphocytes, indicating opioids' ability to
42 modulate the immune system. Liang et al. (2016) stated that chronic use of morphine (at least 12
43 months) causes an increase of μ opioid receptor (MOR) mRNA expression in T lymphocytes (also in B
44 lymphocytes) compared to healthy subjects (Liang et al., 2016). Interestingly, this increase is induced
45 by CD3/CD28-mediated T cell activation, and the transcription of MOR is further mediated by
46 activator protein-1 (AP-1), NF- κ B, and NFAT. These proteins are activated following CD28 ligation
47 (Börner et al., 2008), as mentioned above (Section 2.1).

48

49 Another *in vitro* study demonstrated that morphine inhibits the transcription of IL-2, a hallmark
50 cytokine of T cell activation, as well as AP-1, NF- κ B, and NFAT, which transactivate IL-2. Furthermore,
51 the incubation of opioids (morphine, β -endorphin, and D-penicillamine2-D-penicillamine5-
52 enkephalin) causes a marked increase of cAMP, which in turn activates protein kinase A that
53 augments the activity of C-terminal Src kinase culminating in blocking the initiation of TCR signaling
54 (Börner et al., 2009). Thus, the overexpression of MOR leads to T cell function impairment at the
55 activation stage.

56

57 In the professional APC that links the innate and adaptive immune systems, Wang et al. (2011)
58 showed that IL-23, produced by dendritic cells, was inhibited by morphine in the *S.*
59 *pneumoniae* infection model (Wang et al., 2011). Nevertheless, the disruption of IL-23 is considered
60 in the context of innate immunity. In terms of the APC function in T cell activation, morphine
61 exposure down-regulates MHC class II expression, especially on B cells, which inhibits the activation
62 and proliferation of the CD4⁺ T cell (Roy et al., 2011). Single morphine injection (10 mg/kg) has been
63 shown to decrease basal MHC class II protein expression on B lymphocytes through the
64 hypothalamic-pituitary-adrenal axis. In the central nervous system, opioids activate the
65 hypothalamus, increasing corticosterone release from the adrenal gland. Thus, corticosterone could
66 also reduce basal MHC class II expression. Additionally, this suppression was not presented in
67 morphine-treated adrenalectomized rat models. Eventually, the authors showed that prolonged
68 morphine exposure (using protocol demonstrated to produce tolerance) does not down-regulate
69 MHC-II expression, while morphine withdrawal exerts both a renewed increase in circulating
70 corticosterone levels and a renewed suppression of MHC-II (Nugent et al., 2011; Roy et al., 2011).

71

72 However, not all immune cells are favorable to the antitumor response like the immunosuppressive
73 regulatory T (Treg) cell. Long-term morphine exposure (12 weeks) exhibited an increase of circulating

74 Treg (CD25⁺FoxP3⁺) cells by approximately five-fold in PBMCs sample of rhesus macaques (Cornwell
75 et al., 2013). Furthermore, in breast cancer patients who have undergone resection and were given
76 sufentanil or fentanyl, their Treg number increased after seven days compared to day zero before
77 anesthesia. However, this study did not determine the patients' long-term outcome and the Treg
78 function (Boland and Pockley, 2018; Gong et al., 2014). It is also reported that the percentage of Treg
79 cells obtained from 23 new cases of NSCLC was significantly higher than 16 healthy volunteers.
80 Moreover, this proportion was higher with the increase of cancer stage and in metastatic stage
81 (Erfani et al., 2012), indicating Treg cells' role in cancer progression. Given Treg cells' capacity to
82 repress antitumor immunity, their role becomes crucial for patients with immunotherapy. Takeuchi
83 and Nishikawa (2016) mentioned that Treg cells express immune checkpoint molecules such as CTLA-
84 4 and PD-1. Thus, the success of antibodies blocking these molecules depends on Treg cells'
85 depletion that expresses CTLA-4 or PD-1 (Takeuchi and Nishikawa, 2016).

86
87 Although research has possibly identified how opioids can affect immunity, appropriate *in vitro* and
88 *in vivo* studies must elucidate the detailed mechanism in which the opioids could affect ICI efficacy.
89 Thus, exploring mechanisms could help predict the interaction between opioids and ICIs to avoid
90 detrimental effects when used concomitantly.

91

92 3.2 Opioids directly affect cancer cells

93 Opioids can also directly affect cancer cells due to MOR overexpression in several types of NSCLC.
94 MOR expression on human NSCLC biopsies has revealed a significant increase compared to healthy
95 adjacent tissue samples using immunohistochemistry. Narrowed analysis of samples with lymph
96 node metastasis have demonstrated even a higher increase of MOR than the total lung cancer
97 cohort. Furthermore, MOR staining intensity also increased with the increase of cancer stage
98 (Singleton et al., 2014). Thus, cancer may potentially progress and metastasize through Akt and
99 mTOR pathways. In this case, both exogenous and endogenous opioids can have the same effect,
100 although the former is still debatable (Lennon et al., 2012). Furthermore, morphine has been shown
101 to induce EGFR phosphorylation via MOR, leading to downstream signaling of MAPK/ERK, Akt, and
102 corroborating cancer cell proliferation and invasion. Therefore, there is direct involvement of opioids
103 in the progression of NSCLC (Fujioka et al., 2011). From the other perspective, there is an opportunity
104 to develop adjunctive therapy using antagonists targeting peripheral opioid receptors.

105

106 3.3 Opioid causes gut dysbiosis

107 Morphine and other opioid types interrupt the immune system directly to the immune cell or
108 indirectly to the first layer of surface barriers located in the gut. In the gut lumen, a symbiotic

109 relationship is established between commensal microbiota and the host immunity constructed by
110 intestinal epithelial cells and several mucosal barriers. This complex and dynamic interaction
111 contributes to gut homeostasis, whereby the gut microbiota stimulates epithelial cells, releasing
112 cytokines and chemokines, inducing T cells, and delivering antigens to APC in lymphoid tissue. These
113 barriers also spatially segregate microorganisms and host immune cells avoiding unnecessary
114 immune response. Moreover, intestinal microbiota provides vital aids by digesting dietary fibers,
115 producing vitamin B and K, and metabolizing bile acids (Okumura and Takeda, 2017). Therefore, any
116 changes in the structural or functional microbiota known as “gut dysbiosis” increase the susceptibility
117 to pathogenic infection while reciprocally, the host immune dysregulation causes a perturbation in
118 the intestinal microbiome as well (Cianci et al., 2019; Wang and Roy, 2017).

119

120 Chronic opioid administration has been associated with several GI-adverse reactions, such as
121 constipation, bloating, nausea, and vomiting. Precisely, it further affects (1) intestinal function by
122 inhibiting protective mucus and bicarbonate secretion and disrupting coordination of myenteric
123 activity, hence prolonging the transit time and increasing the risk potential of bacterial translocation,
124 (2) intestinal epithelial integrity by damaging tight junction protein (ZO-1) coordination via activation
125 of TLR (toll-like receptor) 2 and 4 by MOR ligands, and (3) gut microbial by shifting composition
126 towards the expansion of gram-positive pathogens resulting in a reduction of bile-deconjugating
127 bacteria strains (Banerjee et al., 2016; Plein and Rittner, 2018; Wang and Roy, 2017). Thus, opioids
128 possess harmful effects on the gut microenvironment leading to disruption of the immune system.

129

130 3.4 COX inhibitor for NSCLC patient: Analgesic or anticancer?

131 In contrast to opioids, NSAIDs seem to have a favorable effect on the immune system in eradicating
132 tumor cells. A wealth of evidence obtained from *in vitro* and *in vivo* studies has demonstrated that
133 COX-2 is expressed in many solid tumors, including NSCLC, and is associated with tumor proliferation,
134 metastasis, poor prognosis, and resistance to certain antitumor and even radiation (Hattar et al.,
135 2013; Jiang et al., 2019; Wang, 2019; Zhang et al., 2019).

136

137 In 2018, a meta-analysis of seven randomized controlled trials (RCTs) involving 1,559 patients
138 revealed that adding celecoxib to systemic cancer therapy or radiotherapy did not significantly
139 improve the overall response rate (ORR) compared to the placebo. However, further subgroup
140 analysis considering the line of treatment showed that concomitant celecoxib with first-line
141 chemotherapy significantly increased the ORR while the second-line treatment showed no significant
142 difference (Yi et al., 2018). Unfortunately, this study did not consider the expression of COX-2 from
143 tumor cells as not all patients had abundant expression.

144

145 COX-2 expression analysis can be beneficial as a biomarker to determine the prognosis value of the
146 disease or as a predictive marker for patients who may benefit from COX-2 inhibitors. To our
147 knowledge, the latest systematic review and meta-analysis explaining the association between COX-2
148 expression and its impact on patient survival was published in 2013. Two studies were issued almost
149 concurrently, with some similar resources. However, their results were slightly different. Jiang et al.
150 (2013) stated that overexpressed COX-2 in NSCLC patients' tumor biopsies was implicated with the
151 poor prognosis, while Zhan et al. (2013) showed no significant impact. Nonetheless, both research
152 teams had a common result with their subgroup analysis for stage I NSCLC: COX-2 overexpression
153 had a detrimental effect on patient survival (Jiang et al., 2013; Zhan et al., 2013). Thus, at the earliest
154 phase of NSCLC development, COX-2 could serve as either a prognostic factor of the disease or a
155 predictor in selecting patients eligible for adjuvant cancer therapy.

156

157 The first biomarker-driven phase III RCT in the US, CALGB 30203, used the COX-2 index to select
158 patients eligible for the study before they were randomized for the treatment arm receiving
159 celecoxib or the placebo arm. Despite this advanced method, the PFS or OS value of the celecoxib
160 group did not considerably improve compared to the placebo group, including the subgroup of
161 patients whose tumors had COX-2 index ≥ 4 . Hence, the authors argued against using
162 immunohistochemistry (IHC) in examining COX-2, suggesting the use of urinary prostaglandin E2
163 metabolite (PGE-M) instead. PGE-M allows for a real-time measurement compared to IHC, which is
164 usually only assessed once and has an issue in antibodies' specificity (Edelman et al., 2017).

165

166 **4. Drug Interaction between Analgesics and ICIs**

167 The potency of long-term and concomitant exposure to analgesics and ICIs is undeniable. However,
168 the role of analgesic use for relieving pain in cancer patients treated with ICIs lacks clinical evidence.
169 Hence, this review discusses these drugs' possible interactions, drawing connections based on their
170 pharmacodynamic activities. Generally, chronic opioid exposure is associated with the shorter
171 survival of NSCLC patients due to opioids' immunosuppressive properties (Hasegawa et al., 2018;
172 Wang et al., 2015; Zylla et al., 2014). Meanwhile, cancer therapy employing ICI requires a robust
173 immune response from the patient; thus, antitumor like T cells still have enough energy to limit
174 cancer growth and possibly eradicate it. As mentioned, opioids may hamper ICIs within the immune
175 system in many ways, such as reducing T cell function through the overexpression of MOR (Börner et
176 al., 2009, 2008) or downregulation of MHC class II (Liang et al., 2016), increasing immunosuppressor
177 Treg cells (Erfani et al., 2012), directly promoting tumor cell proliferation and invasion (Singleton et
178 al., 2014), and interrupting the gut microbiome composition leading to disruption of gut homeostasis

179 and the whole immune system (Banerjee et al., 2016; Plein and Rittner, 2018; Wang and Roy, 2017;
180 Figure 3).

181

182 Regarding the effect of opioids interrupting ICI through microbiota, Routy et al. (2018) demonstrated
183 that gut microbiome composition influences anti-PD-1 efficacy. They also showed that fecal
184 microbiota transplantation (FMT) taken from ICI-sensitive cancer patients enhanced the antitumor
185 activity of PD-1 inhibitor in germ-free or antibiotic-treated mice. In contrast, FMT from ICI-insensitive
186 cancer patients did not show a significant result. This latter group was then treated with oral
187 supplementation of *Akkermansia muciniphila* (considered “good bacteria”), resulting in the
188 restoration of PD-1 blockade activity (Routy et al., 2018). Furthermore, other studies have explained
189 that ICI (anti-CTLA-4 and PD-1) response requires the presence of distinct bacteria species such as
190 *Bacteroides* sp. and *Bifidobacterium* (Sivan et al., 2015; Vétizou et al., 2015).

191

192 However, to our knowledge, no article explicitly explains the correlation between opioid exposure
193 and ICI efficacy, especially with the case of NSCLC. Therefore, we can only hypothesize that opioids
194 affect ICI efficacy based on two separate articles. For instance, one article stated that opioids cause
195 gut dysbiosis (Banerjee et al., 2016), while another article stated that gut microbiota influences anti-
196 PD-1 efficacy (Routy et al., 2018). Thus, we conclude that opioids impact ICI efficacy through gut
197 homeostasis disruption. Therefore, further *in vitro* and *in vivo* studies are needed to prove our
198 hypothesis.

199

200 Besides, two retrospective studies evaluated the effect of opioids on the prognosis of patients
201 treated with ICIs. The first study from Taniguchi et al. (2020) reported that regular opioid usage
202 during nivolumab treatment initiation impairs the ORR and progression-free survival (PFS) of NSCLC
203 patients receiving nivolumab monotherapy (Taniguchi et al., 2020). Likewise, Iglesias-Santamaria
204 (2020) also showed that concomitant use of opioids had a detrimental effect on the ICI outcome in
205 patients with advanced cancer (including NSCLC) (Iglesias-Santamaría, 2020). Therefore, in the
206 clinical setting, it is suggested to choose opioids with the least effect on immune function
207 deliberately. A list of opioids categorized by the degree of immune modulation effect appears in
208 Table 2. Moreover, dosage regimentation is another critical aspect to consider. Zylla et al. (2014)
209 demonstrated that a group of patients with mild to no pain requiring <5 mg of daily oral morphine
210 equivalent (OME) had a longer survival rate compared to those requiring ≥5 mg/day OME (Zylla et al.,
211 2014). However, Hasegawa et al. (2018), who stratified patients according to daily OME of <60 mg or
212 ≥60 mg, did not indicate a difference in patient survival (Hasegawa et al., 2018). Hence, opioid use
213 for advanced-stage NSCLC patients should not exceed the dose equal to OME of 5 mg per day.

214

215 **Table 2. Opioids categorized by the degree of their immunosuppressive effect** (Zajackowska et al.,
216 2018)

Strong immune modulation	Weak or no immune modulation
Morphine	Buprenorphine (the least or no effect)
Fentanyl	Oxycodone
Sufentanil	Hydromorphone
Codeine	Tramadol

217

218 In contrast, the lower class of analgesics acting on inhibition of the COX-2 enzyme was suggested to
219 support anticancer activity instead of being repressive to the immune system, as shown in an article
220 written by Kobayashi et al. (2020). They reported a case of a 57-years old-male with adenocarcinoma
221 who was treated with nivolumab and celecoxib. As an additional treatment, celecoxib could recover
222 the sensitivity of nivolumab, whereas the patient experienced tumor regrowth after two years
223 following nivolumab monotherapy (Kobayashi et al., 2020). Moreover, a retrospective study
224 evaluating concomitant use of COX inhibitor (aspirin, NSAID, selective COX-2 inhibitor) and ICI
225 revealed that among 37 metastatic NSCLC patients, those receiving the drug combination had
226 significantly longer time-to-progression (TTP) and ORR than ICI alone (Wang et al., 2020). However,
227 Kanai et al. (2021) showed the contrary that COX inhibitor did not have additional or negative
228 impacts on the ICI efficacy based on PFS, OS, response rate, and disease control rate (Kanai et al.,
229 2021). Likewise, COX-2 inhibition in lung cancer cell lines did not affect the PD-L1 expression
230 suggesting no significant benefit of celecoxib during ICI treatment. Although, PD-L1 expression was
231 significantly associated with the COX-2 expression in the resected lung adenocarcinoma specimens
232 (Shimizu et al., 2018). Hence, the usefulness of COX inhibitors is still debatable and necessitates
233 further prospective study (e.g., randomized controlled trial).

234

235 However, in a real situation, for particular patients, NSAIDs are not adequate for relieving moderate-
236 to-severe pain, while opioids as a last resource come with concerning effects. Hence, effective pain
237 management must balance providing pain-free periods without disrupting the causal treatment. For
238 the clinician, the first consideration is the drug of choice. Following WHO and NCCN guidelines to use
239 non-opioids as the first step seems to be the safest way. Next, the clinician may move to opioids with
240 minimum immune modulation (Table 2) before using morphine in response to previous treatment's
241 inadequacy. Lastly, potential adverse events from the combination of analgesics and ICIs should be
242 managed, including immune-related adverse events.

243

244 **5. Conclusion and Future Direction**

245 In conclusion, prolonged opioid use should be limited or replaced with NSAIDs whenever possible to
246 prevent negative impacts on the current immunotherapy and increase the survival rate. Moreover,
247 due to lack of evidence, further studies on concomitant medications using analgesics (either NSAIDs
248 or opioids) and ICIs are essential to avoid unexpected side effects and not compromise both drugs'
249 therapeutic efficacy. Proposed studies should explore the detailed mechanism in which the
250 interaction occurred compared with dosage regimentation, usage duration, drug selection, and
251 administration route.

252

253 **6. Author contributions**

254 RAP conducted the literature search and selection, then wrote the manuscript; MMA designed the
255 outline study, analyzed information, and contributed to manuscript writing; FE contributed to study
256 design and critically reviewed the manuscript. All of the authors have read and approved the final
257 manuscript.

258

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262

263 **8. Declaration of interests**

264 The authors declare no competing interest.

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491 **Figure legends**

492 **Figure 1. Mechanism of CTLA-4 modulating the immune response**

493 The CTLA-4-mediated immune checkpoint is activated in naïve T cells during their initial response to the
494 antigen. Once CD28 is ligated to the ligands (CD80/CD86), it conveys positive or stimulatory signals to the
495 T cell along with signals from the MHC-TCR interaction. Subsequently, the signals triggers CTLA-4 to be
496 expressed on the T cell surface. The stronger the induction through the TCR (and CD28), the greater the
497 amount of CTLA-4 appears on the T cell surface. Hence, the negative signals from CTLA-4 eventually
498 dampens the stimulatory signal to maintain the balance of T cell activation. This figure is adapted from
499 Pardol (2012) (Pardoll, 2012) and created with BioRender.com. TCR, T cell receptor; MHC, major
500 histocompatibility complex; DC, Dendritic cell.

501

502 **Figure 2. Mechanism of PD-1/PD-L1 modulating the immune response**

503 a. In tumor or stromal cells, PDL1 is not constitutively presented but rather is expressed in response to
504 activated T cell inflammatory signals. b. In some tumors, oncogenes constitutive signaling can enhance
505 PD-L1 expression on cancer cells, regardless of inflammatory signals in the tumor microenvironment. This
506 figure is adapted from Pardol (2012)(Pardoll, 2012), created with BioRender.com. MHC, major
507 histocompatibility complex; IFN- γ , interferon- γ ; STATs, signal transducer and activator of transcriptions;
508 TNF- α , tumor nuclear factor- α ; TCR, T cell receptor.

509

510 **Figure 3. Possible pharmacodynamic interactions between ICIs and analgesics (opioids and COX**
511 **inhibitors)**

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